



Partner information:
Not Tested

Physician:
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 ATTN: Strickland Sophie
 Repromed
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Laboratory:
Fulgent Therapeutics LLC
 CAP#: 8042697
 CL A#: 05D2043189
 Laboratory Director:
 Lawrence M Weiss MD
 Report Date: **May 17, 2024**

Access on:
FT-6969909

Test #: FT-TS14824643
 Specimen Type: Saliva Swab
 Collected: No Provided

Access on:
N/A

FINAL RESULTS



Carr er for **ONE** genet c cond t on
 Genet c counse ng s
 recommended.

TEST PERFORMED

Monash Beacon Expanded Male Carrier Screening Panel v2.1

(363 Gene Panel; gene sequencing with deletion and
 duplication analysis)

Condition and Gene	Inheritance	Partner
Congenital adrenal hyperplasia due to 21-hydroxylase deficiency CYP21A2	AR	N/A

+ Poss b e Carr er
 c.955C>T(*) 2C>T +
 CYP21A2 duplication
 p.(Gln39*)()

INTERPRETATION:

Notes and Recommendations:

- Based on these results, this individual is positive for a carrier mutation in 1 gene. Carrier screening for the reproductive partner is recommended to accurately assess the risk for any autosomal recessive conditions. A negative result reduces, but does not eliminate, the chance to be a carrier for any condition included in this screen. Please see the supplemental table for details.
- Testing for copy number changes in the SMN1 gene was performed to screen for the carrier status of Spina Muscular Atrophy. The results for this individual are within the normal range for non-carriers. See Limitations section for more information.
- This carrier screening test does not screen for all possible genetic conditions, nor for all possible mutations in every gene tested. This report does not include variants of uncertain significance; only variants classified as pathogenic or likely pathogenic at the time of testing, and considered relevant for reproductive carrier screening, are reported. Please see the gene specific notes for details. Please note that the classification of variants can change over time.
- Patients may wish to discuss any carrier results with their doctor, as there is an increased chance that they are also carriers. These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- X-linked genes are not routinely analyzed for male carrier screening tests. Gene specific notes and mutations may be present. See below.
- Genetic counseling is recommended. Contact your physician about the available options for genetic counseling.



CONGENITAL ADRENAL HYPERPLASIA DUE TO 21-HYDROXYLASE DEFICIENCY

Patient	Partner
Result	N/A
Variant Details	N/A

What is Congenital adrenal hyperplasia due to 21-hydroxylase deficiency?

Congenital adrenal hyperplasia (CAH) due to 21-hydroxylase deficiency is an inherited disorder that affects the adrenal glands and hormone production. Approximately 75 percent of individuals with classic 21-hydroxylase deficiency have the salt-wasting type, whereby the body excretes too much salt in urine. Affected infants present with poor feeding, weight loss, dehydration, and vomiting, all of which can be life-threatening. Females with this condition typically have ambiguous genitalia, while males usually have normal genitalia, but with small testes. Individuals with the simple virilizing form and the non-classic form of the disease do not experience salt loss. Males and females with either the classic or non-classic forms of 21-hydroxylase deficiency tend to have an early growth spurt, but the final adult height is usually shorter than others in the family, and affected individuals may have reduced fertility. Additionally, individuals may have excessive body hair growth, hair loss, and irregular menstruation. Some individuals (male or female) with the non-classic form of the disease may have mild, non-life-threatening symptoms, while others may never develop symptoms of the disorder at all.

What is my risk of having an affected child?

CAH due to 21-hydroxylase deficiency is inherited in an autosomal recessive manner. If the patient and the partner are both carriers, the risk for an affected child is 1 in 4 (25%).

What kind of medical management is available?

Treatment consists of early initiation of hormone replacement therapy and/or surgery for females. Prognosis is good for patients with appropriate medical management and psychological support.

What mutation was detected?

The heterozygous variants c.955C>T (p.G n319*) and a whole gene duplication of CYP21A2 were detected in this sample. In addition, the benign polymorphism c.*12C>T was also detected. The phase of these variants is unknown but could be determined through parental testing.

The nonsense variant, p.G n319*, introduces a premature stop codon and is expected to result in the loss of function of the protein product of the CYP21A2 gene, either as the result of protein truncation or of nonsense-mediated mRNA decay. This variant, also reported as Q318*, is a classic 21-hydroxylase-deficient congenital adrenal hyperplasia mutation and has been reported in multiple affected individuals (PubMed: 3267225, 12220458, 12915679). The variant, p.G n319*, and the polymorphism c.*12C>T are known to frequently occur in a duplicated copy of the CYP21A2 gene coexisting with a normal copy of CYP21A2 on the same chromosome. This haplotype was identified in approximately 2% of the general population and in ~80% of carriers of p.G n319*, and such a configuration may represent a benign allele (PubMed: 28401898, 19773403). Nonetheless, there is a possibility that p.G n319* occurs on a chromosome with only a single copy of CYP21A2, in which case it results in a pathogenic allele. If multiple copies of CYP21A2 are present, we cannot be certain if the p.G n319* variant occurs on a chromosome with one (i.e. pathogenic state) or two (i.e. benign state) copies of CYP21A2. While this combination of variants may represent a benign allele, the laboratory classifies the variant p.G n319* as a key pathogenic.



GENES TESTED:

Monash Beacon Expanded Male Carrier Screening Panel v2.1 - 363 Genes

This analysis was run using the Monash Beacon Expanded Male Carrier Screening Panel v2.1 gene set. 363 genes were tested with 99.5% of targets sequenced at >20x coverage. For more gene-specific information and assistance with results, see the SUPPLEMENTAL TABLE.

ABCA12 ABCA3 ABCA4 ABCB11 ABCC8 ACAD9 ACADM ACADVL ACAT1 ACOX1 ACSF3 ADA ADAMTS2 ADGRG1 ADK AGA AGL AGPS AGXT AHI1 AIPL1 ALDH3A2 ALDOB ALG6 ALMS1 ALPL AMT AQP2 ARG1 ARL13B ARSA ARSB ASL ASNS ASPA ASS1 ATM ATP6V1B1 ATP7B BBS1 BBS10 BBS12 BBS2 BCKDHA BCKDHB BCS1L BLM BSND CAPN3 CASQ2 CBS CC2D2A CCDC103 CCDC39 CCDC88C CDH23 CEP290 CFTR CHRNE CHRNG CHST6 CIITA CLN3 CLN5 CLN6 CLN8 CLRN1 CNGB3 COL27A1 COL4A3 COL4A4 COL7A1 COX15 CPS1 CPT1A CPT2 CRB1 CRYL1 CTNS CTSA CTSC CTSD CTSK CYBA CYP11A1 CYP11B1 CYP11B2 CYP17A1 CYP1B1 CYP21A2 CYP27A1 DBT DCLRE1C DDX11 DHCR7 DHDDS DLD DNAH5 DNAI1 DNAI2 DUOX2 DUOX2A DYNC2H1 DYSF EIF2AK3 EIF2B5 ELP1 ERCC2 ERCC5 ERCC6 ERCC8 ESCO2 ETFA ETFB ETFDH ETHE1 EVC EVC2 EXOSC3 F2 F5 FAH FAM126A FAM161A FANCA FANCC FANCG FH FKRP FKTN FOXRED1 FTCD FUCA1 G6PC GAA GALT GALNS GALT GAMT GBA GBE1 GCDH GDAP1 GDF5 GFM1 GJB2 GJB6 GLB1 GLDC GLE1 GNE GNPTAB GNPTG GNS GSS GUCY2D GUSB HADHA HADHB HAX1 HBA1 HBA2 HBB HEXA HEXB HGSNAT HJV HLCS HMGCL HOGA1 HPS1 HPS3 HPS4 HSD17B4 HSD3B2 HYLS1 IDUA IVD IYD JAK3 KCNJ11 LAMA2 LAMA3 LAMB3 LAMC2 LCA5 LDLRAP1 LHX3 LIFR LIPA LMBRD1 LOXHD1 LPL LRP2 LRPPRC LYST MAN2B1 MANBA MCOLN1 MCPH1 MED17 MESP2 MFSDB MKS1 MLC1 MLYCD MMAA MMAB MMACHC MMADHC MPI MPL MPV17 MTHFR MTMR2 MTRR MTTP MUT MVK MYO7A NAGA NAGLU NAGS NBN NDRG1 NDUFAF2 NDUFAF5 NDUFS4 NDUFS6 NDUFS7 NDUFV1 NEB NEU1 NPC1 NPC2 NPHP1 NPHS1 NPHS2 NTRK1 OAT OCA2 OPA3 OTOF P3H1 PAH PANK2 PC PCCA PCCB PCDH15 PCNT PDHB PEX1 PEX10 PEX12 PEX2 PEX26 PEX6 PEX7 PFKM PHGDH PHYH PKHD1 PLA2G6 PLOD1 PMM2 POLG POLR1C POMGNT1 POMT1 POMT2 POR PPT1 PRF1 PROP1 PSAP PTS PUS1 QDPR RAB23 RAG1 RAG2 RAPS N RARS2 RAX RDH12 RMRP RNASEH2B RPE65 RPGRIP1L RTKL1 SACS SAMD9 SAMHD1 SCO2 SEPSECS SERPINA1 SGCA SGCB SGCD SGCG SGSH SH3TC2 SLC12A6 SLC17A5 SLC19A3 SLC1A4 SLC22A5 SLC25A13 SLC25A15 SLC26A2 SLC26A3 SLC35A3 SLC37A4 SLC39A4 SLC45A2 SLC46A1 SLC5A5 SLC7A7 SMARCAL1 SMN1 SMPD1 SPG11 SPINK5 STAR SUMF1 SURF1 TCIRG1 TCTN2 TECPR2 TF TG TGM1 TH TMEM216 TPO TPP1 TRDN TRIM32 TRMU TSEN54 TSFM TSHB TTC37 TTPA TYMP TYR TYR1 UGT1A1 USH1C USH1G USH2A VPS13A VPS13B VPS45 VPS53 VPK1 VSX2 WHRN WRN XPA XPC ZFYVE26

METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 99.54% and 99.49% of coding regions and splicing junctions of genes tested had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus-specific databases, literature searches, and other molecular biology computational pipelines. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions (+/- 20bp) of the genes tested above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("full"). A gene tested were evaluated for deletions and/or duplications. However, single exons or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications are analyzed using Fulgent Genome proprietary pipeline for this specimen. Bioinformatics: The Fulgent Genome v2019.2 pipeline was used to analyze this specimen.

LIMITATIONS:

General Limitations

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated family relationships, and use of the correct human reference sequences at the queried locus. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or otherwise, to future pregnancies, and negative results do not rule out the genetic risk to a pregnancy. Off-catalog gene names change over time. Fulgent uses the most up-to-date gene names based on HUGO Gene Nomenclature Committee (<https://www.genenames.org>) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and a manual annotation available at the time of reporting. This assay is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions other than specified genes. DNA alterations in regulatory



regions or deep intron regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm, however these types of alterations are not detected as readily as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. A though next generation sequencing technologies and our bioinformatics analysis software may reduce the confounding contrast but not of pseudogene sequences or other highly homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analyses can identify alterations of genomic regions which include one whole gene (buccal swab specimens and whole blood specimens) and are two or more contiguous exons in size (whole blood specimens only); single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analyses has not been performed on regions that have been sequenced by Sanger.

Gene Specific Notes and Limitations

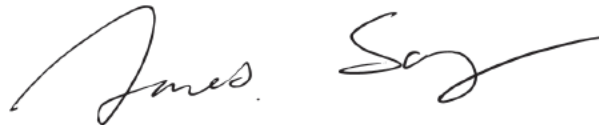
CEP290: Copy number analysis for exons 8-13 and exons 39-42 may have reduced sensitivity in the CEP290 gene. Confirmation of these exons are mediated via a positive personal history of CEP290-related conditions and/or individuals carrying a pathogenic/ key pathogenic sequence variant. **CFTR:** Analysis of the non-8 polymorphic region (e.g. IVS8-5T allele) is only performed if the p.Arg117 (R117) mutation is detected. Single exon deletion/duplication analysis is limited to deletions of previously reported exons: 1, 2, 3, 11, 19, 20, 21. CFTR variants primarily associated with CFTR-related so-called congenital absence of the vas deferens and CFTR-related pancreatitis are not included in this analysis. CFTR variant with insufficient evidence of being cystic fibrosis mutations will not be reported either. **CRYL1:** As mutations in the CRYL1 gene are not known to be associated with any clinical condition, sequence variants in this gene are not analyzed. However, an increase copy number detected for large deletions including this gene and a neighboring gene on the same (GJB6, also known as connexin 30), this gene was evaluated for copy number variation. **CYP11B1:** The current sequencing method is not able to reliably detect certain pathogenic variants in this gene due to the interference by highly homologous regions. This analysis is not designed to detect or rule-out copy-number changes in CYP11B1/CYP11B2 gene. **CYP11B2:** The current sequencing method is not able to reliably detect certain pathogenic variants in this gene due to the interference by highly homologous regions. This analysis is not designed to detect or rule-out copy-number changes in CYP11B1/CYP11B2 gene. **CYP21A2:** Significant pseudogene interference and/or reciprocal exchanges between the CYP21A2 gene and its pseudogene, CYP21A1P, have been known to occur and may impact results. As such, the relevance of variant reported in this gene must be interpreted in the context of the clinical findings, biochemical profile, and family history of each patient. CYP21A2 variant primarily associated with non-classical congenital adrenal hyperplasia (CA) are not included in this analysis (PubMed: 23359698). The variant associated with non-classical disease, including but not limited to c.188A>T (p. S63Leu), c.844G>T (p. Val282Leu), c.1174G>A (p. A392Thr), and c.1360C>T (p. Pro454Ser) will not be reported. LR-PCR is not routinely ordered for NM 000500.9:c.955C>T (p. G319Ter). Individual with c.955C>T (p. G319Ter) will be reported as a Possible Carrier indicating that the precise nature of the variant has not been determined by LR-PCR and that the variant may occur in the CYP21A2 wild-type gene or in the CYP21A1P pseudogene. The confirmation is recommended for the second reproductive partner is tested positive for variant associated with classical CA. **DDX11:** Due to the interference by highly homologous regions, our current sequencing method has less sensitivity to detect variants in the DDX11 gene. **DUOX2:** The current sequencing method is not able to reliably detect variants in exons 6-8 of the DUOX2 gene (NM 014080.5) due to significant interference by the highly homologous gene, DUOX1. **F2:** The common risk allele NM 000506.5:c.*97G>A is not included in this analysis. **F5:** The common Factor 5 "Leiden" allele is not typically reported as this variant is associated with low disease penetrance. **GALT:** In general, the D2 "Duar e" allele is not reported if detected, but can be reported upon request. While this allele can cause positive newborn screening results, is not known to cause clinical symptoms in any state (PubMed: 25473725, 30593450). **GBA:** The current sequencing method may not be able to reliably detect certain pathogenic variants in the GBA gene due to homologous recombination between the pseudogene and the functional gene. **HBA1:** Significant interference from highly homologous regions in exons 1-2 of the BA1 gene has been recognized to occur, potentially impeding the assay's technical capability to detect pathogenic alterations during sequencing analyses. **HBA2:** Significant interference from highly homologous regions in exons 1-2 of the BA2 gene has been recognized to occur, potentially impeding the assay's technical capability to detect pathogenic alterations during sequencing analyses. **HSD17B4:** Copy number analysis for exons 4-6 may have reduced sensitivity in the SD17B4 gene. Confirmation of these exons are mediated via a positive personal history of D-b functional protein deficiency and Perrault syndrome and/or individuals carrying a pathogenic/ key pathogenic sequence variant. **LMBRD1:** Copy number analysis for exons 9-12 may have reduced sensitivity in the LMBRD1 gene. Confirmation of these exons are mediated via a positive personal history of combined meyloma and homocystinuria and/or individuals carrying a pathogenic/ key pathogenic sequence variant. **MTHFR:** As recommended by ACMG, the two common polymorphisms in the MTHFR gene - c.1286A>C (p. G429A, also known as c.1298A>C) and c.665C>T (p. A222Val, also known as c.677C>T) - are not reported in this test due to lack of sufficient clinical utility over the sequencing (PubMed: 23288205). **NEB:** This gene contains a 32-kb repeat region (exons 82-105) which





is not amenable to sequencing and deletion/duplication analysis. NPHS2: If detected, the variant NM_014625.3:c.686G>A (p.Arg229Gln) will not be reported as this variant is not significantly associated with disease when homozygous or in the compound heterozygous state with variants in exons 1-6 of NP_52. SERPINA1: If detected the variant NM_000295.5:c.863A>T (p.Glu288Val) will not be reported as this variant is associated with low disease penetrance and is not associated with severe early onset disease. SMN1: The current sequencing method detects sequencing variants in exon 7 and copy number variations in exons 7-8 of the SMN1 gene (NM_022874.2). Sequencing and deletion/duplication analysis are not performed on any other region of this gene. About 5%-8% of the population have two copies of SMN1 on a single chromosome and a deletion on the other chromosome, known as a [2+0] configuration (PubMed: 20301526). The current sequencing method cannot detect carriers with a [2+0] SMN1 configuration, but can detect linkage between the sex carrier allele and certain population-specific single nucleotide changes. As a result, a negative result for carriers sequencing greatly reduces but does not eliminate the chance that a person is a carrier. Only abnormal results will be reported. TRDN: Due to high GC content of certain exons (including exons 4-5), copy number analysis may have reduced sensitivity for paralogous deletions/duplications of TRDN. Confirmation of paralogous deletions/duplications are mediated via duals with a positive personal history of cardiac arrhythmia and/or duals carrying a pathogenic/likely pathogenic sequence variant. TYR: Due to the interference by highly homozygous regions, our current sequencing method has less sensitivity to detect variants in exons 4-5 of the TYR gene (NM_000372.5). UGT1A1: Common variants in the UGT1A1 gene (population allele frequency >5%) are typically not reported as they do not cause a Mendelian condition. VPS45: LoF is not a known disease mechanism. WRN: Due to the interference by highly homozygous regions within the WRN gene, our current sequencing method has less sensitivity to detect variants in exons 10-11 of WRN (NM_000553.6).

SIGNATURE:



Jianbo Song, Ph.D., ABMGG, CGMB, CCS, FACMG on 5/17/2024
Laboratory Director, Fulgent

DISCLAIMER:

This test was developed and its performance characteristics determined by **Fulgent Therapeutics LLC**. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high-complexity testing. This test is used for clinical purposes. It should not be regarded as investment or for research. Since genetic variation, as well as systematic and technical factors, can affect the accuracy of testing, the results of testing should always be interpreted in the context of clinical and family data. For assistance with interpretation of these results, healthcare professionals may contact us directly at (626) 350-0537 or info@fulgentgenetics.com. It is recommended that patients receive appropriate genetic counseling to explain the implications of the test results, including risks, uncertainties and reproductive or medical options.

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To view the supplemental table describing the carrier frequencies, detection rates, and residual risks associated with the genes on this test please visit the following link:

[Beacon Expanded Carrier Screening Supplemental Table](#)

